

1. Sella L., Govind R., Caracciolo R., Quarantin A., Vu V.V., Tundo S., Nguyen H.M., Favaron F., Musetti R., De Zotti M. (2021). Transcriptomic and ultrastructural analyses of *Pyricularia oryzae* treated with fungicidal peptaibol analogs of *Trichoderma trichogin*. *Frontiers in Microbiology*, 12:753202. DOI: 10.3389/fmicb.2021.753202.
2. Tundo S., Paccanaro M.C., Bigini V., Savatin D.V., Faoro F., Favaron F., Sella L. (2021). The *Fusarium graminearum* FGSG_03624 xylanase enhances plant immunity and increases resistance against bacterial and fungal pathogens. *International Journal of Molecular Sciences*, 22(19): 10811. DOI:org/10.3390/ijms221910811.
3. De Zotti M.^a, Sella L.^a, Bolzonello A., Gabbatore L., Peggion C., Bortolotto A., Elmaghraby I., Tundo S. and Favaron F. (2020). Targeted amino acid substitutions in *Trichoderma* peptaibol confer activity against fungal plant pathogens and protect host tissues from *Botrytis cinerea* infection. *International Journal of Molecular Sciences*, 21: 7521. DOI:10.3390/ijms21207521. (^aAuthors equally contributing to the work)
4. Echeverrigaray S., Scariot F.J., Fontanella G., Favaron F., Sella L., Santos M.C., Schwambach J., Pedrotti C., Delamare A.P.L. (2020). *Colletotrichum* species causing grape ripe rot disease in *Vitis labrusca* and *V. vinifera* varieties in the highlands of southern Brazil. *Plant Pathology*, 69: 1504-1512. DOI: dx.doi.org/10.1111/ppa.13240.
5. Tundo S., Paccanaro M.C., Elmaghraby I., Moscetti I., D'Ovidio R., Favaron F., Sella L. (2020). The xylanase inhibitor TAXI-I increases plant resistance to *Botrytis cinerea* by inhibiting the BcXyn11a xylanase necrotizing activity. *Plants*, 9 (5): 601. DOI:10.3390/plants9050601.
6. Linaldeddu B.T., Bregant C., Montecchio L., Favaron F., Sella L.* (2020). First report of *Phytophthora acerina*, *P. pini* and *P. plurivora* causing root rot and sudden death of olive trees in Italy. *Plant Disease*, doi: 10.1094/PDIS-10-19-2080-PDN (*corresponding author)
7. Luti S.*, Sella L.*, Quarantin A., Pazzagli L., Baccelli I. (2020). Twenty years of research on cerato-platanin family proteins: clues, conclusions, and unsolved issues. *Fungal Biology Reviews*, 34: 13-24. Doi: 10.1016/j.fbr.2019.10.001 (*authors contributing equally to the work)
8. Quarantin A., Haderl B., Kröger B., Schaefer W., Favaron F., Sella L.*, Martinez-Rocha A.L. (2019). Different hydrophobins of *Fusarium graminearum* are involved in hyphal growth, attachment, water-air interface penetration and plant infection. *Frontiers in Microbiology*. DOI: 10.3389/fmicb.2019.00751. (*corresponding author)
9. Quarantin A., Castiglioni C., Schäfer W., Favaron F., Sella L.* (2019). The *Fusarium graminearum* cerato-platanins loosen cellulose substrates enhancing fungal cellulase activity as expansin-like proteins. *Plant Physiology and Biochemistry*, 139: 229–238. DOI: 10.1016/j.plaphy.2019.03.025. (*corresponding author)
10. Heidari B., Miras M., Maria B., Lucini L., Bolton M., Mcgrath M.J., Broccanello C., Alberti I., Sella L., Concheri G., Squartini A., Cagnin M., Hassani M., Romano A., Stevanato P. (2019). Mass spectrometry-based metabolomic discrimination of *Cercospora* leaf spot resistant and susceptible sugar beet gemplasms. *Euphytica* 2019. DOI: 10.1007/s10681-019-2351-3.
11. Zanella A., Ascher-Jenuil J., Ponge J.F., Bolzonella C., Banas D., De Nobili M., Fusaro S., Sella L., Giannini R. (2018). Humusica: Soil biodiversity and global change. *Bulletin of Geography-Physical Geography Series*, 14: 15-36. DOI: 10.2478/bgeo-2018-0002.
12. Hanson L., De Lucchi C., Stevanato P., McGrath M., Panella L., Sella L., De Biaggi M., Concheri G. (2018). Root rot symptoms in sugar beet lines caused by *Fusarium oxysporum* f. s.p. *betae*. *European Journal of Plant Pathology*, 150: 589-593. DOI 10.1007/s10658-017-1302-x.
13. Dal Cortivo C., Conselvan G.B., Carletti P., Barion G., Sella L., Vamerali T. (2017). Biostimulant Effects of Seed-Applied Sedaxane Fungicide: Morphological and Physiological Changes in Maize Seedlings. *Frontiers in Plant Science*, 8, 2072. doi: 10.3389/fpls.2017.02072.
14. Paccanaro M.C., Sella L.*, Castiglioni C., Giacomello F., Martinez-Rocha A.L., D'Ovidio R., Schäfer W., Favaron F. (2017). Synergistic Effect of Different Plant Cell Wall-Degrading Enzymes Is Important for Virulence of *Fusarium graminearum*. *Molecular Plant-Microbe Interactions*, 30 (11): 886-895. (*corresponding author)
15. De Lucchi C., Stevanato P., Hanson L.E., McGrath J.M., Panella L., De Biaggi M., Broccanello C., Bertaglia M., Sella L., Concheri G. (2017). Molecular markers for improving control of soil-borne pathogen *Fusarium oxysporum* in sugar beet. *Euphytica*, 213:71.
16. Quarantin A., Glasenapp A., Schäfer W., Favaron F., Sella L.* (2016). Involvement of the *Fusarium graminearum* cerato-platanin proteins in fungal growth and plant infection. *Plant Physiology and Biochemistry*, 109: 220-229. (*corresponding author)
17. Marcato R., Sella L., Lucchetta M., Vincenzi S., Odorizzi S., Curioni A., Favaron F. (2016). Necrotrophic fungal plant pathogens display different mechanisms to counteract grape chitinase and thaumatin-like protein. *Physiological and Molecular Plant Pathology*, doi: 10.1016/j.pmpp.2016.09.005.
18. Sella L.*, Castiglioni C., Paccanaro M.C., Janni M., Schäfer W., D'Ovidio R., Favaron F. (2016). Involvement of fungal pectin methyl esterase activity in the interaction between *Fusarium graminearum* and wheat. *Molecular Plant-Microbe Interactions*, 29 (4): 258-267 (doi: 10.1094/MPMI-07-15-0174-R). (*corresponding author)
19. Sella L.*, Gazzetti K., Castiglioni C., Schäfer W., D'Ovidio R., Favaron F. (2016). The *Fusarium*

- graminearum Xyr1 transcription factor regulates xylanase expression but is not essential for fungal virulence. *Plant Pathology*, 65 (5): 713-722 (doi: 10.1111/ppa.12456). (*corresponding author)
20. Tacconi, G., Paltrinieri, S., Mejia, J.F., Fuentealba, S.P., Bertaccini, A., Tosi, L., Giacomini, A., Mazzucchi, U., Favaron, F., Sella, L., Bertaiola, F. (2015). Vine decline in kiwifruit: climate change and effect on waterlogging and Phytophthora in North Italy. *Acta Horticulturae*, 1096: 93-98 (doi: 10.17660/ActaHortic.2015.1096.7).
 21. Tundo S., Moschetti I., Faoro F., Lafond M., Giardina T., Favaron F., Sella L., D'Ovidio R. (2015). *Fusarium graminearum* produces different xylanases causing host cell death that is prevented by the xylanase inhibitors XIP-I and TAXI-III in wheat. *Plant Science*, 240: 161-169.
 22. Moschetti I., Faoro F., Moro S., Sabbadin D., Sella L., Favaron F., D'Ovidio R. (2015). The xylanase inhibitor TAXI-III counteracts the necrotic activity of a *Fusarium graminearum* xylanase in vitro and in durum wheat transgenic plants. *Molecular Plant Pathology*, 16: 583-592.
 23. Kalunke R.M., Cenci A., Volpi C., O'Sullivan D.M., Sella L., Favaron F., Cervone F., De Lorenzo G., D'Ovidio R. (2014). The pgip family in soybean and three other legume species: evidence for a birth-and-death model of evolution. *BMC Plant Biology*, 14: 189. DOI: 10.1186/s12870-014-0189-3.
 24. Sella L., Gazzetti K., Castiglioni C., Schäfer W., Favaron F. (2014). *Fusarium graminearum* possesses virulence factors common to *Fusarium* head blight of wheat and seedling rot of soybean, but differing in their impact on disease severity. *Phytopathology*, 104: 1201-1207.
 25. Moschetti I., Tundo S., Janni M., Sella L., Gazzetti K., Tauzin A., Giardina T., Masci S., Favaron F., D'Ovidio R. (2013). Constitutive expression of the xylanase inhibitor TAXI-III delays *Fusarium* Head Blight symptoms in durum wheat transgenic plants. *Molecular Plant-Microbe Interactions*, 26: 1464-1472.
 26. Sella L.*, Gazzetti K., Faoro F., Odorizzi S., D'Ovidio R., Schäfer W., Favaron F. (2013). A *Fusarium graminearum* xylanase expressed during wheat infection is a necrotizing factor but is not essential for virulence. *Plant Physiology and Biochemistry*, 64: 1-10. (*corresponding author)
 27. Scattolin L., Dal Maso E., Mutto Accordi S., Sella L., Montecchio L. (2012). Detecting asymptomatic ink-diseased chestnut trees by the composition of the ectomycorrhizal community. *Forest Pathology*, 42: 501-509.
 28. Ferrari S.a, Sella L.a, Janni M.a, De Lorenzo G., Favaron F., D'Ovidio R. (2012). Transgenic expression of Polygalacturonase-Inhibiting Proteins in *Arabidopsis* and wheat increases resistance to the flower pathogen *Fusarium graminearum*. *Plant Biology*, 14 (Supplement 1): 31-38. (a Authors equally contributing to the work).
 29. Kalunke R.M., Janni M., Sella L., David P., Geffroy V., Favaron F., D'Ovidio R. (2011). Transcript analysis of the bean polygalacturonase inhibiting protein gene family reveals that Pvgip2 is expressed in the whole plant and is strongly induced by pathogen infection. *Journal of Plant Pathology*, 93: 141-148.
 30. Sella L., Cosmi T., Giacomello F., Saccardi A., Favaron F. (2010). First report of *Fusarium oxysporum* on *Dipladenia* sp. in Italy. *Journal of Plant Pathology*, 92: 543-543.
 31. Favaron F., Lucchetta M., Odorizzi S., Da Cunha A.T.P. and Sella L. (2009). The role of grape polyphenols on trans-resveratrol activity against *Botrytis cinerea* and of fungal laccase on the solubility of putative grape PR proteins. *Journal of Plant Pathology*, 91: 579-588.
 32. Tomassini A., Sella L., Raiola A., D'Ovidio R. and Favaron F. (2009). Characterization and expression of *Fusarium graminearum* endo-polygalacturonases in vitro and during wheat infection. *Plant Pathology*, 58: 556-564.
 33. Janni M., Sella L., Favaron F., Blechl A.E., De Lorenzo G., D'Ovidio R. (2008). The expression of a bean PGIP in transgenic wheat confers increased resistance to the fungal pathogen *Bipolaris sorokiniana*. *Molecular Plant-Microbe Interactions*, 21: 171-177.
 34. Raiola A., Sella L., Castiglioni C., Balmas V., Favaron F. (2008). A single amino acid substitution in highly similar endo-PGs from *Fusarium verticillioides* and related *Fusarium* species affects PGIP inhibition. *Fungal Genetics and Biology*, 45: 776-789.
 35. Mosca E., Montecchio L., Sella L., Garbaye J. (2007). Short-term effect of removing tree competition on the ectomycorrhizal status of a declining pedunculate oak forest (*Quercus robur* L.). *Forest Ecology and Management*, 244: 129-140.
 36. D'Ovidio R., Roberti S., Di Giovanni M., Capodicasa C., Melaragni M., Sella L., Tosi P., Favaron F. (2006). The characterization of the soybean polygalacturonase inhibiting proteins (Pgip) gene family reveals that a single member is responsible for the activity detected in soybean tissue. *Planta*, 224: 633-645.
 37. Sella L., Tomassini A., D'Ovidio R., Favaron F. (2005). Expression of two *Sclerotinia sclerotiorum* endo-pg genes correlates with endo-polygalacturonase activity during *Glycine max* infection. *Journal of Plant Pathology*, 87: 199-205.
 38. Zuppini A., Navazio L., Sella L., Castiglioni C., Favaron F., Mariani P. (2005). An endopolygalacturonase from *Sclerotinia sclerotiorum* induces a Ca²⁺-mediated signalling and programmed cell death in soybean cells. *Molecular Plant-Microbe Interactions*, 18: 849-855.
 39. Sella L., Castiglioni C., Roberti S., D'Ovidio R., Favaron F. (2004). An endo-polygalacturonase (PG) of *Fusarium moniliforme* escaping inhibition by plant polygalacturonase-inhibiting proteins

(PGIPs) provides new insights into the PG-PGIP interaction. *FEMS Microbiology Letters*, 240: 117-124.

40. Favaron F., Sella L., D'Ovidio R. (2004). Relationships among endo-polygalacturonase, oxalate, pH and plant polygalacturonase-inhibiting protein (PGIP) in the interaction between *Sclerotinia sclerotiorum* and soybean. *Molecular Plant-Microbe Interactions*, 17: 1402-1419.

Capitoli di libro

1. Sella L., V.V. Vu, A. Quarantin, R. Caracciolo, R. Govind, A. Bolzonello, S. Tundo, M. De Zotti, F. Favaron, H.D. Nguyen, Q.L. Le, T.T. Nguyen, L.T. Do, H.M. Nguyen (2021). Sustainable methods to control *Pyricularia oryzae*, the causal agent of rice blast disease. In: Anderle M. (eds) *Innovations in Land, Water and Energy for Vietnam's Sustainable Development*, pp 67-82. UNIPASpringer Series. Springer, Cham. https://doi.org/10.1007/978-3-030-51260-6_7. First Online: 29 November 2020. Print ISBN: 978-3-030-51259-0. Online ISBN: 978-3-030-51260-6.

Pubblicazioni brevi su riviste internazionali

1. De Zotti M., Sella L., Formaggio F., Favaron F. (2018). Peptide-Based Biopesticides. Meeting Abstract: OP5. *Journal of Peptide Science*, 24: S49.
2. De Zotti M., Sella L., Quarantin A., Castiglioni C., Bolzonello A., Favaron F., Picco A.M., Govind R., Tundo S., Odorizzi S., Le T.D., Vu V.V., Nguyen M.H. (2018). Novel peptide-based control measures against the rice fungal pathogen *Pyricularia oryzae*. Meeting Abstract: P178. *Journal of Peptide Science*, 24: S144.
3. Sella L. (2005). Fungal pathogens can elude the inhibition by host plant polygalacturonase inhibiting proteins (PGIPs) by different mechanisms. *Journal of Plant Pathology*, 87 (special issue 4): 265.

Pubblicazioni per esteso in proceedings di congressi

1. Tacconi G., Tosi L., Giacomini A., Mazzucchi U., Favaron F., Sella L., Bertaiola F., Mejia J.F., Paltrinieri S., Fuentealba S.P., Bertaccini A. (2014). Moria dell'actinidia nel veronese: anomalie climatiche, struttura del terreno e ruolo dei patogeni. Convegno della Società di Ortoflorofruitticoltura Italiana (SOI). Latina, 25-11-2014.
2. D'Ovidio R., Roberti S., Melaragni M., Capodicasa C., Sella L., Favaron F. (2002). Characterization of two closely linked soybean pgip genes and transcript regulation following pathogen infection and wounding. *Proceedings of 6th Congress of the European Foundation for Plant Pathology. Plant Protection Science*, 38 (Special issue 2): 480-482.
3. Favaron F., D'Ovidio R., Melaragni M., Sella L., Destro T. (2000). PGIP activities and PGIP gene family structure in soybean and leek. *Proceedings of the 5th Congress of the European Foundation for Plant Pathology*, pp. 346-348.